ABSTRACT.—Infectious diseases can potentially affect the seasonal migration of hawks. We tested three species of California hawks from autumn 2004 to spring 2006 for haemopodid blood parasites (Plasmodium, Haemoproteus, and Leucocytozoon spp.). We screened 323 Red-tailed Hawks (Buteo jamaicensis), 100 Cooper’s Hawks (Accipiter cooperii), and 21 Red-shouldered Hawks (B. lineatus) for blood parasites using polymerase chain reaction (PCR). Among these hawks, 221 (50%) were infected with Haemoproteus, 134 (30%) with Leucocytozoon, and none with Plasmodium species. We compared blood parasite prevalence results to identify infection patterns in two populations (birds captured during autumn at a migration watchsite in the Marin Headlands [Golden Gate Raptor Observatory] vs. birds captured during winter in the Central Valley) and between years, seasonal periods, and sex and age classes. In 2004, Marin Red-tailed Hawks were more likely to test positive for blood parasites if sampled late in the migration; however, we observed the opposite pattern in Cooper’s Hawks. The prevalence of both Haemoproteus and Leucocytozoon followed the same seasonal trends in each host species, which suggested a possible effect of infection status on raptor migration. We also observed different patterns of variation in parasite prevalence between years at the two sampling sites, but no significant differences between juvenile and adult birds at either site. In addition, using a simple measure of body condition, we observed significant differences between males and females with regard to the effects of parasite infection and sampling location. These results provide a baseline for future studies monitoring long-term changes in parasite prevalence and effects on California’s migrating hawks.

KEY WORDS: Cooper’s Hawk; Accipiter cooperii; Red-tailed Hawk; Buteo jamaicensis; blood parasites; migration.
Blood parasites are commonly found in birds and can have serious consequences for their hosts during energy-demanding periods such as migration and reproduction (Remple 2004, Garvin et al. 2006). Migratory birds often have higher prevalences of blood parasites than nonmigrants (Møller and Erritzøe 1998). This has been attributed to the stress of migration and the fact that migrants move through more terrain, which increases the likelihood of encountering a higher diversity of parasite vectors (Møller and Erritzøe 1998, Smith et al. 2004, Valkiunas 2005). Although migration occurs in both spring and autumn, most previous parasite studies have focused on spring migration due to the increased prevalence of haematozoan infections in spring and summer (Allander and Bennett 1995, Phalen et al. 1995, Rintamäki et al. 1998, 1999, Dawson and Bortolotti 2000, Dyracz et al. 2005, Garvin et al. 2006). Few studies have examined blood parasites during autumn and winter months when parasite infections likely become latent (Schrader et al. 2003, Garvin et al. 2004, Smith et al. 2004).

Migration requires a significant energy investment and is an especially treacherous time for juvenile birds (Clarabuch and Gonzáles-Solís 1998). Hawks, in particular, have high first-year mortality rates. Band recovery data suggest that first-year mortality rates can range from 50–83% (Newton 1979). Although most blood parasites are thought to be nonpathogenic, parasite-induced fatalities can occur within a bird’s first year of life (Davidar and Morton 1993, Leppert et al. 2004). Parasite infections acquired by nestlings presumably could negatively affect both the timing of migration departure and the bird’s ability to survive (Hunter et al. 1997, Leppert et al. 2004). Thus, blood-parasite studies during autumn and winter may reveal parasite effects on migrating juvenile hawks.

We studied blood-parasite prevalences in three hawk species (Accipitridae) sampled in California. We examined Red-tailed Hawks (Buteo jamaicensis), Red-shouldered Hawks (B. lineatus), and Cooper’s Hawks (Accipiter cooperii) captured at an autumn migration count site in the Marin Headlands on the central coast, and wintering Red-tailed Hawks and Red-shouldered Hawks captured in the Central Valley. Taking seasonality into consideration, we examined differences in blood parasite prevalence between the two study populations and between sex and age classes.

**METHODS**

**Sample Collection.** We collected blood samples from hawks captured during autumn at the Golden Gate Raptor Observatory (GGRO) migration watchsite located in the Marin Headlands, Sausalito, CA (37°40’N, 122°20’W) and from a dense concentration of wintering hawks in the northern Central Valley of California (38°05’–39°57’N, 121°22’–122°17’W). We captured hawks at the GGRO site between 15 August and 19 December in 2004 and 2005 using bow nets, dho-gaza traps, and mist nets (Bloom et al. 2007). We captured wintering raptors using bal-chatri traps along roadsides (Berger and Mueller 1959) from 26 November 2004 to 6 March 2005 and from 25 November 2005 to 22 February 2006. In this study, the 2004 season refers to all birds captured from August 2004 through March 2005, and the 2005 season encompasses all captures from August 2005 through February 2006.

We banded each captured hawk with a U.S. Geological Survey aluminum leg band, identified each to species and approximate age, and measured mass and wing chord for each. We determined the sex of Red-tailed Hawks as described by Pitzer et al. (2008). We also collected a few drops of blood from the medial metatarsal vein of each bird. We used one drop of blood to make a blood smear fixed with methanol and stained with Giemsa (Godfrey et al. 1987). We placed the rest of the blood sample in 1 ml of lysis buffer (Sehgal et al. 2006) and stored it in a −20°C freezer. All captured hawks appeared healthy and were released at the trap site.

**Parasite Detection.** We extracted parasite DNA and screened blood samples for *Leucocytozoon* spp. following methods described by Sehgal et al. (2006).
To test for *Plasmodium* and *Haemoproteus* spp., we used a second PCR protocol that amplifies the cytochrome *b* region of the mtDNA. For this PCR reaction we used the same PCR components from Sehgal et al. (2006) with the following primers: L15183 and H15730 (Fallon et al. 2003, Szymanski and Lovette 2005). This PCR reaction does not distinguish between the two parasite genera. The cycling profile is similar to Sehgal et al. (2006) with a denaturing step for 50 sec and an annealing temperature of 53°C for 50 sec.

We used positive and negative controls to detect *Plasmodium*, *Haemoproteus*, and *Leucocytozoon*. Positive controls were from birds with known infections evident from microscopy results, and we used water for the negative controls. We visualized the PCR products on 2% agarose gels stained with ethidium bromide under UV light.

To distinguish between the *Plasmodium* and *Haemoproteus* genera, we sequenced DNA from 88 hawks (64 Red-tailed Hawks and 24 Cooper’s Hawks) using the L15183 and H15730 primers. We performed bidirectional sequencing with dye-terminator fluorescent labeling in an ABI Prism 3100 automated sequencer (Applied Biosystems, Inc., Foster City, California, U.S.A.). We sequenced 512 base pairs and edited them using Sequencher 3.1 (GeneCodes, Ann Arbor, Michigan, U.S.A.). We then identified sequences to genus by identifying their closest sequence matches in GenBank via the National Center for Biotechnology Information (NCBI) nucleotide BLAST search. The most similar sequences were all *Haemoproteus* spp. with the following accession numbers: AF465592, AF46591, and AF465593.

We conducted phylogenetic analyses using maximum parsimony techniques and PAUP* 4.0b10 (Swofford 2002). Searches used the bootstrap search option with 1000 stepwise addition replicates and the TBR branch-swapping algorithm. We constructed a simple consensus tree to summarize the results and estimated genetic divergences using the HKY85 setting. We deposited all sequences in GenBank (FJ966919 to FJ966927).

We examined some Giemsa-stained slides for haemosporidian parasites using a Nikon E200 compound microscope at 100×, 400×, and 1000×.

**Statistical Tests.** We first used a one-way chi-square test to evaluate overall differences among species in prevalences of the two parasite genera (*Haemoproteus* sp. and *Leucocytozoon* sp.). We then used multiple logistic regression to determine if there were significant differences within host species in the prevalence (binary response variable: infected or not) of different blood parasites (*Haemoproteus* and *Leucocytozoon* analyzed separately) between years (2004 vs. 2005), age groups (juvenile vs. adult), regions (Marin Headlands vs. Central Valley) and date of capture (before and after 15 October). We ran these models for Red-tailed Hawks and Cooper’s Hawks, but not for Red-shouldered Hawks due to small sample sizes. The Red-tailed Hawk model included all variables plus all relevant two- and three-way interactions, whereas the Cooper’s Hawk model included only year, age, and date of capture, because we sampled this species only in the Marin Headlands. We tested the association between infection status and predictor variables using generalized linear models (Proc GENMOD with binomial distribution of errors and logit link function; SAS Institute 1999).

We also investigated relationships between body condition and the occurrence of blood parasites. We considered only Red-tailed Hawks for these analyses due to small sample sizes and incomplete sets of mass and wing-length data for Cooper’s Hawks and Red-shouldered Hawks. We calculated an index of body condition as measured by residuals from an ordinary least squares (OLS) regression of mass against a flat wing measurement (Schulte-Hostedde et al. 2005). Red-tailed Hawks exhibit sexual dimorphism, wherein females tend to be heavier than males, although substantial overlap exists (Pitzer et al. 2008). To account for the effect of sex on our body condition indices, we calculated the OLS residuals individually for each sex. We then ran an ANOVA to test for relationships between body-condition indices and explanatory variables: one infection-status variable for each parasite (binary; yes or no), sex (male vs. female), year (2004 vs. 2005), age (juvenile vs. adult), region (Marin Headlands vs. Central Valley), date of capture (before and after 15 October), and relevant two-way interactions.

**Results**

We collected blood samples from 323 Red-tailed Hawks (Marin Headlands: *n* = 183; Central Valley: *n* = 140), 100 Cooper’s Hawks (Marin Headlands: *n* = 100), and 21 Red-shouldered Hawks (Marin Headlands: *n* = 6; Central Valley: *n* = 15). Among these, 134 of 446 (30%) tested positive for *Leucocytozoon* and 221 (49.5%) tested positive for *Haemoproteus/Plasmodium* infections. Sequencing of parasite mtDNA from a subset of hawks (88) revealed eight lineages of *Haemoproteus* infections, but no cas-
es of Plasmodium infection. In Red-tailed Hawks, the species visible was Haemoproteus elani, but blood smears were of insufficient quality to identify species in Cooper’s Hawks (G. Valkiuunas unpubl. data). Maximum likelihood and HKY85 distance analyses identified two Haemoproteus clades with a 4% average sequence divergence (Fig. 1). One clade was specific to Cooper’s Hawk parasites with intraspecific variation of 1.9%, and a second clade consisted of mostly Red-tailed Hawk parasites with intraspecific sequence divergence of 0.3%.

Overall blood-parasite prevalence was highest in Cooper’s Hawks (61.2% Leucocytozoon, 49.8% Haemoproteus) and lowest in Red-shouldered Hawks (9.5% Leucocytozoon, 14.3% Haemoproteus; Table 1). Chi-square analyses confirmed significant overall host species differences between Red-tailed Hawks and Cooper’s Hawks in the prevalence of both parasite groups (Leucocytozoon: $\chi^2 = 69.46$, $P < 0.001$; Haemoproteus: $\chi^2 = 12.23$, $P = 0.002$). The small sample size for Red-shouldered Hawks did not allow us to run the statistical model for this species.

The logistic regression model for Cooper’s Hawks and Haemoproteus infections indicated a significant year effect, with more infected birds in 2004 than in 2005 ($\chi^2 = 26.41$, $P < 0.001$), and a significant date effect, with more individuals infected before 15 October than later in the migration season ($\chi^2 = 21.71$, $P < 0.001$). For Leucocytozoon infections, we found no significant year effect; however, Cooper’s Hawks also showed more Leucocytozoon infections early in the migration season ($\chi^2 = 4.92$, $P < 0.026$). No significant ($P \leq 0.05$) age effects or interactions were evident in either model.

Table 1. Prevalence (% infected) of Haemoproteus (H) and Leucocytozoon (L) in Red-tailed Hawks, Cooper’s Hawks, and Red-shouldered Hawks sampled during autumn migration in the Marin Headlands and during winter in the Central Valley of California.

<table>
<thead>
<tr>
<th>SPECIES</th>
<th>LOCATION</th>
<th>YEAR</th>
<th>AGE</th>
<th>N</th>
<th>H</th>
<th>L</th>
</tr>
</thead>
<tbody>
<tr>
<td>Red-tailed Hawk</td>
<td>Marin Headlands</td>
<td>2004</td>
<td>Juvenile</td>
<td>143</td>
<td>67</td>
<td>29</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>9</td>
<td>78</td>
<td>22</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2005</td>
<td>Juvenile</td>
<td>28</td>
<td>29</td>
<td>29</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>3</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Central Valley</td>
<td>2004</td>
<td>Juvenile</td>
<td>60</td>
<td>47</td>
<td>8</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>47</td>
<td>43</td>
<td>11</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2005</td>
<td>Juvenile</td>
<td>27</td>
<td>22</td>
<td>26</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>6</td>
<td>33</td>
<td>17</td>
</tr>
<tr>
<td>Cooper’s Hawk</td>
<td>Marin Headlands</td>
<td>2004</td>
<td>Juvenile</td>
<td>29</td>
<td>90</td>
<td>66</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>2</td>
<td>100</td>
<td>5</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2005</td>
<td>Juvenile</td>
<td>66</td>
<td>33</td>
<td>65</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>5</td>
<td>20</td>
<td>6</td>
</tr>
<tr>
<td>Red-shouldered Hawk</td>
<td>Marin Headlands</td>
<td>2004</td>
<td>Juvenile</td>
<td>4</td>
<td>25</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2005</td>
<td>Juvenile</td>
<td>2</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Central Valley</td>
<td>2004</td>
<td>Juvenile</td>
<td>5</td>
<td>20</td>
<td>40</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>4</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2005</td>
<td>Juvenile</td>
<td>2</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Adult</td>
<td>4</td>
<td>25</td>
<td>0</td>
</tr>
</tbody>
</table>
For Red-tailed Hawks and *Haemoproteus* prevalence, the logistic regression model indicated significant interactions between year and site ($\chi^2 = 7.11$, $P = 0.007$) and between year and date ($\chi^2 = 10.24$, $P = 0.0014$). The first interaction term indicated higher infection rates at the Marin Headlands in 2004 than in 2005, whereas Central Valley birds were similarly infected in both years (Fig. 2). The second interaction term indicated that Red-tailed Hawks captured after 15 October during the 2004 migration season were significantly more likely to be infected with *Haemoproteus* than those captured before 15 October in the same season; however, the opposite was true in 2005 (Fig. 3). No other significant main effects or interactions were evident in either model.

For Red-tailed Hawks and *Leucocytozoon* prevalence, we again found significant interactions between year and site ($\chi^2 = 6.89$, df = 1, $P = 0.008$) and between year and date ($\chi^2 = 5.28$, $P = 0.022$). In this case, infection prevalence did not differ between years in the Marin Headlands, whereas individuals were more infected in 2005 than in 2004 in the Central Valley (Fig. 2). Interestingly, though, we found the same seasonal trend for *Leucocytozoon* infections as for *Haemoproteus* (Fig. 3).

The ANOVA model of Red-tailed Hawk body condition indicated significant main effects for sex ($F_{1,272} = 44.88$, $P < 0.001$), site ($F_{1,272} = 23.99$, $P < 0.001$), and *Haemoproteus* infection status ($F_{1,272} = 5.16$, $P = 0.024$), as well as significant interactions between sex and site ($F_{1,272} = 14.12$, $P < 0.001$) and between sex and *Haemoproteus* infection status ($F_{1,272} = 6.06$, $P = 0.014$). Given the strong significance of sex as a main effect and two relevant interaction terms, we also ran separate sex-specific analyses with all other explanatory variables and relevant interactions included in each model. For female Red-tailed Hawks, we found only a site effect, which indicated that females captured in the Marin Headlands were in poorer condition than those captured in the Central Valley ($F_{1,118} = 33.38$, $P < 0.001$). For males, we did not find a site effect ($F_{1,153} = 0.75$, $P = 0.38$), but males captured before 15 October were in better condition than those captured later in the season ($F_{1,153} = 6.61$, $P = 0.011$). In addition, males with a *Haemoproteus* infection were in poorer condition than those without such an infection ($F_{1,153} = 13.35$, $P < 0.001$). We saw no evidence of a change in body condition with *Leucocytozoon* infection, between years, or between age groups.

**Discussion**

Overall, 50% of the hawks from this study were infected with *Haemoproteus* and 30% harbored *Leucocytozoon* infections. Although we sequenced only a random subset of individuals, our results suggested that the prevalence of *Plasmodium* infections was likely absent or very low. We did not sequence every individual for which a positive *Haemoproteus* or Plas-
modium infection was indicated, because we believe the extent of variation was adequately confirmed by the fact that 54 individuals showed the same Haemoproteus lineage. Greiner et al. (1975) and Ziman et al. (2004) also recorded high prevalences of Haemoproteus and Leucocytozoon via microscopy, but no Plasmodium parasites in diurnal raptors on the West Coast.

We found that collection year accounted for significant differences in the prevalence of Haemoproteus among Red-tailed and Cooper’s hawks (Table 1). For both species, Haemoproteus prevalence was much higher in 2004 than in 2005, and among Central Valley Red-tailed Hawks, Leucocytozoon prevalence was higher in 2005 than in 2004. Other studies have noted yearly differences in parasite prevalence and have attributed this temporal variation to variation in vector abundance (Greiner et al. 1975, Van Riper et al. 1986, Sol et al. 2000, Scheuerlein and Ricklefs 2004) and numbers of potential host species (Ricklefs et al. 2005).

We suspect, however, that the significant differences in Haemoproteus prevalence between years most likely are related to the conspicuously different capture rates for Red-tailed Hawks in the Marin Headlands in the two years. In 2005, GGRO volunteers captured only 12 Red-tail Hawks after 15 October, compared to 109 captured during the same period the previous year, and the overall capture rate for Red-tailed Hawks was unusually low that year (n = 121, including hawks not tested for blood parasites) when compared to the mean capture rate of 303 per yr (data from 22 yr of collection; GGRO unpubl. data). The reason for the reduced number of Red-tailed Hawks in 2005 is unknown and may reflect a variety of factors beyond the scope of this study. Due to the unusually low number of birds captured in 2005, we propose that future research hypotheses stemming from this work should be based on only the 2004 data.

Within the subset of Red-tailed Hawks from the Marin Headlands, we found that the prevalence of Haemoproteus and Leucocytozoon increased significantly during the second half of the 2004 (but not 2005) migration season (Fig. 3). We observed the opposite pattern (significantly more infected before 15 Oct) for both parasite species among Cooper’s Hawks. Variation in blood-parasite prevalence between early and late migrants has been documented in other avian studies. For example, Rintamäki et al. (1998) found a pattern in Willow Warblers (Phylloscopus trochilus), in which individuals infected with Haemoproteus migrated earlier than those infected with Leucocytozoon. That study also documented a possible trend in which individuals infected with Haemoproteus were captured early and the number of infected individuals decreased with time, whereas the number of Leucocytozoon infections increased with time and was highest at the end of migration. The study by Rintamäki et al. (1998) showed differential tim-
ing in the migration of birds infected with the two blood parasites, whereas in our study high infection rates for the two parasites occurred during the same period; i.e., either before or after 15 October, depending on the year and/or hawk species.

Phalen et al. (1995) showed that Sharp-shinned Hawks (*Accipiter striatus*) and Cooper’s Hawks caught late in spring migration had higher parasite prevalences than individuals captured earlier. The authors attributed this phenomenon to spring relapse, which is a seasonal increase in parasite intensity that occurs at the onset of breeding, perhaps initiated by hormone changes and the extra energy burdens of reproduction (Phalen et al. 1995, Schrader et al. 2003). However, we observed the same phenomenon during autumn migration among Red-tailed Hawks, when there is no reproductive stress to induce a spring relapse. Therefore, one hypothesis for the results seen in 2004 is that, among Red-tailed Hawks, the stress of migration alone may be sufficient to weaken the immune system enough to allow parasite infections to persist or worsen during autumn, perhaps causing the infected individuals to migrate behind healthier unparasitized birds. We did not observe the same pattern in Cooper’s Hawks, however. Hence, an alternative explanation may be that, both within and among species, the migrant birds comprise individuals from different source areas that became infected at different locations. Indeed, a recent study by Hull et al. (2009) showed that GGRO migrants caught from 15 August to 30 September appeared to be local dispersers, whereas those caught from 1 October to 30 November most likely included a mixture of birds from California and the Intermountain West. It is also known that there are significant genetic differences among populations of Red-tailed Hawks (Hull et al. 2008). Thus, it is possible that the higher prevalence of *Haemoproteus* in late-season migrants may be attributed to their different sites of origin.

The breeding grounds of the birds may not necessarily be the site of parasite transmission, however. Hasselquist et al. (2007) found that transmission of *Haemoproteus* occurred on migratory stopover sites and wintering grounds, such that the vector location drove the observed blood-parasite prevalence. Manwell (1955) also found completely different rates of infection among eastern and western American Robins (*Turdus migratorius*), which he believed to be a function of differing vector distributions. Accordingly, further research to confirm the source-population dynamics of Marin Headlands migrants and associated parasite transmission vectors may help explain the patterns we observed.

We found no significant differences in parasite prevalence between juveniles and adults of the three species analyzed. Higher parasite prevalence in adults has been recorded in some avian-parasite studies (Davidar and Morton 1993, Garvin and Greiner 2003), but not in others (Rintamäki et al. 1998, Garvin et al. 2003, Ricklefs et al. 2005). Although we detected no age-related differences in parasite prevalence, we were unable to examine parasite intensity (parasitemia). Typically, juveniles experience a more severe acute-phase infection than adults, because they have not had time to develop an acquired immunity (Hunter et al. 1997, Dawson and Bortolotti et al. 2000, Sol et al. 2003, Stjernman et al. 2004). Thus, analyzing only differences in prevalence data between age classes may underestimate the relative effects of blood parasites on juveniles and adults (Sol et al. 2003, Leppert et al. 2004).

Finally, we found that female Red-tailed Hawks in the Marin Headlands averaged significantly lower body condition index scores than those in the Central Valley (also see Pitzer et al. 2008). Although body condition scores between sampling sites could be explained by numerous factors, we assume that hawks with a lower mass to wing length residual score from the same sample site is an indication of poorer health (Hull et al. 2006). Several studies have reported effects of parasitism on body condition indices, and indeed we did find a significant effect in migrating males (Dawson and Bortolotti et al. 2000, Schrader et al. 2003, Stjernman et al. 2004, Dyrsez et al. 2005). Males tested prior to 15 October were in better condition than those captured after 15 October, suggesting that early migrants have better health. Due to the sex discrepancy, however, it is difficult to generalize concerning how these parasites may affect their hosts. We also acknowledge that using residuals from a mass-wing length regression as an index of body condition is a relatively coarse tool for revealing the complexities of physiological interactions between parasites and the body condition of their hosts (Green 2001, Schulte-Hos tedde et al. 2005), which may have precluded our uncovering relatively subtle effects.

In conclusion, our study encompasses the largest sample to date of migratory and wintering hawks tested for blood parasites. Our data provide a baseline that will aid future researchers seeking to monitor changes in infectious diseases in hawk populations, which may arise due to factors such as global
climate change and increasing habitat loss in western North America.

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